

UV Dose Required to Achieve Incremental Log Inactivation of Bacteria, Protozoa and Viruses¹

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1. This compilation has been prepared for Trojan Technologies Inc. and is published here as a public service to the UV community.
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BRIEF DESCRIPTION AND SELECTION CRITERIA FOR CONTENT OF THE TABLES

Tables 1-4 present a summary of published data on the Ultraviolet (UV) dose-response of various organisms that are pathogens, indicators, or organisms encountered in the application, testing of performance, and validation of UV disinfection technologies. The tables reflect the state of knowledge, but include the variation in technique and biological response that currently exists in the absence of standardized protocols. Users of the data for their own purposes are advised to exercise critical judgment in how they use the data.

In most cases, the data are generated from low pressure (LP) monochromatic mercury arc lamp sources for which the lamp fluence rate (intensity) can be measured empirically and multiplied by exposure time to obtain a dose. Earlier data do not always contain the correction factors that are now considered standard practice (Bolton and Linden 2003). Some data are from polychromatic medium pressure (MP) mercury arc lamps, and in some cases both lamp types are used. In a few cases, filtered polychromatic UV light is used to achieve a narrow band of irradiation around 254 nm. These studies are also designated as LP.

None of the data incorporate any impact of photorepair processes. Only the response to the inactivating UV dose is documented. The references from which the data are abstracted must be carefully read to understand how the reported doses are calculated and what the assumptions and procedures are in the calculation.

At the time this table was being prepared, a parallel initiative (Hijnen et al. 2006) was ongoing and is recommended to the reader.

It is the intention of Trojan Technologies, École Polytechnique de Montreal and INRS- Institut Armand-Frappier to keep this table dynamic, with periodic updates. Recommendations for inclusion in the tables, along with the reference source, can be sent to:

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The selection criteria for inclusion are recommended as follows:

1. Data must be already published in a peer-reviewed journal or other peer-reviewed publication media;
2. The dose-response should be empirically determined in the laboratory with the assistance of a collimated beam apparatus;
3. Ideally, the fluence rate (intensity) should be measured with a recently calibrated radiometer and when this has not been done, a well-characterized organism should be run as a reference to provide a comparison with the literature values to substantiate that the radiometer is within calibration.
4. The publication from which the data is abstracted should describe the experimental procedures including collimated beam procedures, dose calculation procedures along with any assumptions made, organism culturing procedures, enumeration and preparation for experiments.
5. Responses should be determined over a range of doses; that is, a complete dose-response curve is preferred to a single dose-response measurement.

Table 1. UV Doses for Multiple Log Reductions for Various Spores

Spore	Lamp Type	UV Dose (Fluence) (mJ/cm ²) for a given Log Reduction without photo-reactivation							Reference
		1	2	3	4	5	6	7	
<i>Bacillus subtilis</i> ATCC6633	N/A	36	48.6	61	78				Chang et al. 1985
<i>Bacillus subtilis</i> ATCC6633	LP	24	35	47	79				Mamane-Gravetz and Linden 2004
<i>Bacillus subtilis</i> ATCC6633	LP	22	38	>50					Sommer et al. 1998
<i>Bacillus subtilis</i> ATCC6633	LP	20	39	60	81				Sommer et al. 1999
<i>Bacillus subtilis</i> WN626	LP	0.4	0.9	1.3	2				Marshall et al., 2003

Table 2. UV Doses for Multiple Log Reductions for Various Bacteria

Bacterium	Lamp Type	UV Dose (Fluence) (mJ/cm ²) for a given Log Reduction without photo-reactivation							Reference
		1	2	3	4	5	6	7	
<i>Aeromonas hydrophila</i> ATCC7966	LP	1.1	2.6	3.9	5	6.7	8.6		Wilson et al. 1992
<i>Aeromonas salmonicida</i>	LP	1.5	2.7	3.1	5.9				Liltved and Landfald 1996
<i>Campylobacter jejuni</i> ATCC 43429	LP	1.6	3.4	4	4.6	5.9			Wilson et al. 1992
<i>Citrobacter diversus</i>	LP	5	7	9	11.5	13			Giese and Darby 2000
<i>Citrobacter freundii</i>	LP	5	9	13					Giese and Darby 2000
<i>Escherichia coli</i> ATCC 11229	N/A	2.5	3	3.5	5	10	15		Harris et al. 1987
<i>Escherichia coli</i> ATCC 11229	N/A	3	4.8	6.7	8.4	10.5			Chang et al. 1985
<i>Escherichia coli</i> ATCC 11229	LP	<5	5.5	6.5	7.7	10			Zimmer et al. 2002
<i>Escherichia coli</i> ATCC 11229	MP	<3	<3	<3	<3	8			Zimmer et al. 2002
<i>Escherichia coli</i> ATCC 11229	LP	7	8	9	11	12			Hoyer 1998
<i>Escherichia coli</i> ATCC 11229	LP	3.5	4.7	5.5	6.5	7.5	9.6		Sommer et al. 2000
<i>Escherichia coli</i> ATCC 11229	LP	6	6.5	7	8	9	10		Sommer et al. 1998
<i>Escherichia coli</i> ATCC 11303	LP	4	6	9	10	13	15	19	Wu et al. 2005
<i>Escherichia coli</i> ATCC 25922	LP	6	6.5	7	8	9	10		Sommer et al. 1998
<i>Escherichia coli</i> C	LP	2	3	4	5.6	6.5	8	10.7	Otaki et al. 2003
<i>Escherichia coli</i> O157:H7	LP	1.5	3	4.5	6				Tosa and Hirata 1999
<i>Escherichia coli</i> O157:H7	LP	<2	<2	2.5	4	8	17		Yaun et al. 2003
<i>Escherichia coli</i> O157:H7 CCUG 29193	LP	3.5	4.7	5.5	7				Sommer et al. 2000
<i>Escherichia coli</i> O157:H7 CCUG 29197	LP	2.5	3	4.6	5	5.5			Sommer et al. 2000
<i>Escherichia coli</i> O157:H7 CCUG 29199	LP	0.4	0.7	1	1.1	1.3	1.4		Sommer et al. 2000
<i>Escherichia coli</i> O157:H7 ATCC 43894	LP	1.5	2.8	4.1	5.6	6.8			Wilson et al. 1992
<i>Escherichia coli</i> O25:K98:NM	LP	5	7.5	9	10	11.5			Sommer et al. 2000
<i>Escherichia coli</i> O26	LP	5.4	8	10.5	12.8				Tosa and Hirata 1999
<i>Escherichia coli</i> O50:H7	LP	2.5	3	3.5	4.5	5	6		Sommer et al. 2000
<i>Escherichia coli</i> O78:H11	LP	4	5	5.5	6	7			Sommer et al. 2000
<i>Escherichia coli</i> K-12 IFO3301	LP & MP	2	4	6	7	8.5			Oguma et al. 2002
<i>Escherichia coli</i> K-12 IFO3301	LP & MP	2.2	4.4	6.7	8.9	11.0			Oguma et al. 2004
<i>Escherichia coli</i> K-12 IFO3301	LP	1.5	2	3.5	4.2	5.5	6.2		Otaki et al. 2003
<i>Escherichia coli</i> Wild type	LP	4.4	6.2	7.3	8.1	9.2			Sommer et al. 1998

Table 2. (continued)

Bacterium	Lamp Type	UV Dose (Fluence) (mJ/cm ²) for a given Log Reduction without photo-reactivation							Reference
		1	2	3	4	5	6	7	
<i>Halobacterium elongata</i> ATCC33173	LP	0.4	0.7	1					Martin et al. 2000
<i>Halobacterium salinarum</i> ATCC43214	LP	12	15	17.5	20				Martin et al. 2000
<i>Klebsiella pneumoniae</i>	LP	12	15	17.5	20				Giese and Darby 2000
<i>Klebsiella terrigena</i> ATCC33257	LP	4.6	6.7	8.9	11				Wilson et al. 1992
<i>Legionella pneumophila</i> ATCC 43660	LP	3.1	5	6.9	9.4				Wilson et al. 1992
<i>Legionella pneumophila</i> ATCC33152	LP	1.6	3.2	4.8	6.4	8.0			Oguma et al. 2004
<i>Legionella pneumophila</i> ATCC33152	MP	1.9	3.8	5.8	7.7	9.6			Oguma et al. 2004
<i>Pseudomonas stutzeri</i>	UVB	100	150	195	230				Joux et al. 1999
RB2256	UVB	175	>300						Joux et al. 1999
<i>Salmonella spp.</i>	LP	<2	2	3.5	7	14	29		Yaun et al. 2003
<i>Salmonella anatum</i> (from human feces)	N/A	7.5	12	15					Tosa and Hirata 1998
<i>Salmonella derby</i> (from human feces)	N/A	3.5	7.5						Tosa and Hirata 1998
<i>Salmonella enteritidis</i> (from human feces)	N/A	5	7	9	10				Tosa and Hirata 1998
<i>Salmonella infantis</i> (from human feces)	N/A	2	4	6					Tosa and Hirata 1998
<i>Salmonella typhi</i> ATCC 19430	LP	1.8	4.8	6.4	8.2				Wilson et al. 1992
<i>Salmonella typhi</i> ATCC 6539	N/A	2.7	4.1	5.5	7.1	8.5			Chang et al. 1985
<i>Salmonella typhimurium</i> (from human feces)	N/A	2	3.5	5	9				Tosa and Hirata 1998
<i>Salmonella typhimurium</i> (from human feces)	N/A	2	3.5	5	9				Tosa and Hirata 1998
<i>Salmonella typhimurium</i> (in act. sludge)	LP	3	11.5	22	50				Maya et al. 2003
<i>Salmonella typhimurium</i>	UVB	50	100	175	210	250			Joux et al. 1999
<i>Shigella dysenteriae</i> ATCC29027	LP	0.5	1.2	2	3	4	5.1		Wilson et al. 1992
<i>Shigella sonnei</i> ATCC9290	N/A	3.2	4.9	6.5	8.2				Chang et al. 1985
<i>Staphylococcus aureus</i> ATCC25923	N/A	3.9	5.4	6.5	10.4				Chang et al. 1985
<i>Streptococcus faecalis</i> ATCC29212	N/A	6.6	8.8	9.9	11.2				Chang et al. 1985
<i>Streptococcus faecalis</i> (secondary effluent)	N/A	5.5	6.5	8	9	12			Harris et al. 1987
<i>Vibrio anguillarum</i>	LP	0.5	1.2	1.5	2				Liltved and Landfald 1996
<i>Vibrio cholerae</i> ATCC25872	LP	0.8	1.4	2.2	2.9	3.6	4.3		Wilson et al. 1992
<i>Vibrio natriegens</i>	UVB	37.5	75	100	130	150			Joux et al. 1999
<i>Yersinia enterocolitica</i> ATCC27729	LP	1.7	2.8	3.7	4.6				Wilson et al. 1992
<i>Yersinia ruckeri</i>	LP	1	2	3	5				Liltved and Landfald 1996

Table 3. UV Doses for Multiple Log Reductions for Various Protozoa

Protozoan	Lamp Type	UV Dose (Fluence) (mJ/cm ²) for a given Log Reduction without photo-reactivation							Reference
		1	2	3	4	5	6	7	
<i>Cryptosporidium hominis</i>	LP & MP	3	5.8						Johnson et al. 2005
<i>Cryptosporidium parvum</i> , oocysts, tissue culture assay	N/A	1.3	2.3	3.2					Shin et al. 2000
<i>Cryptosporidium parvum</i>	LP & MP	2.4	<5	5.2	9.5				Craik et al. 2001
<i>Cryptosporidium parvum</i>	MP	<5	<5	<5	~6				Amoah et al. 2005
<i>Cryptosporidium parvum</i>	MP	<10	<10	<10					Belosevic et al. 2001
<i>Cryptosporidium parvum</i>	LP	1	2	<5					Shin et al. 2001
<i>Cryptosporidium parvum</i>	MP	1	2	2.9	4				Bukhari et al. 2004
<i>Cryptosporidium parvum</i>	LP	<2	<2	<2	<4	<10			Clancy et al. 2004
<i>Cryptosporidium parvum</i>	MP	<3	<3	3-9	<11				Clancy et al. 2000
<i>Cryptosporidium parvum</i>	LP	<3	<3	3-6	<16				Clancy et al. 2000
<i>Cryptosporidium parvum</i>	LP	0.5	1	1.4	2.2				Morita et al. 2002
<i>Cryptosporidium parvum</i>	LP	2	<3	<3					Zimmer et al. 2003
<i>Cryptosporidium parvum</i>	MP	<1	<1	<1					Zimmer et al. 2003
<i>Encephalitozoon cuniculi</i> , microsporidia	LP	4	9	13					Marshall et al. 2003
<i>Encephalitozoon hellem</i> , microsporidia	LP	8	12	18					Marshall et al. 2003
<i>Encephalitozoon intestinalis</i> , microsporidia	LP & MP	<3	3	<6	6				Huffman et al. 2002
<i>Encephalitozoon intestinalis</i> , microsporidia	LP	3	5	6					Marshall et al. 2003
<i>Giardia lamblia</i> , gerbil infectivity assay	LP	<0.5	<0.5	<0.5	<1				Linden et al. 2002b
<i>Giardia lamblia</i>	LP	<10	~10	<20					Campbell et al. 2002
<i>Giardia lamblia</i>	LP	<2	<2	<4					Mofidi et al. 2002
<i>Giardia lamblia</i> , excystation assay	N/A	> 63							Rice and Hoff 1981
<i>Giardia lamblia</i> , excystation assay	N/A	40	180						Karanis et al. 1992
<i>Giardia muris</i> , excystation assay	N/A	77	110						Carlson et al. 1985
<i>G. muris</i> , cysts, mouse infectivity assay	N/A	<2	<6	10 + tailing					Craik et al. 2000
<i>Giardia muris</i>	MP	1	4.5	28 + tailing					Craik et al. 2000
<i>Giardia muris</i>	MP	<10	<10	<25	~60				Belosevic et al. 2001
<i>Giardia muris</i>	LP	<1.9	<1.9	~2	~2.3				Hayes et al. 2003
<i>Giardia muris</i>	LP	<2	<2	<4					Mofidi et al. 2002
<i>G. muris</i> , cysts	MP	<5	<5	5					Amoah et al. 2005

Table 4. UV Doses for Multiple Log Reductions for Various Viruses

Virus	Host	Lamp Type	UV Dose (Fluence) (mJ/cm ²) per Log Reduction						Reference
			1	2	3	4	5	6	
PRD-1 (Phage)	<i>S. typhimurium</i> Lt2	N/A	9.9	17.2	23.5	30.1			Meng and Gerba 1996
B40-8 (Phage)	<i>B. Fragilis</i>	LP	11	17	23	29	35	41	Sommer et al. 2001
B40-8 (Phage)	<i>B. fragilis</i> HSP-40	LP	12	18	23	28			Sommer et al 1998
MS2 (Phage)	<i>Salmonella typhimurium</i> WG49	N/A	16.3	35	57	83	114	152	Nieuwstad and Havelaar 1994

Table 4. (continued)

Virus	Host	Lamp Type	UV Dose (Fluence) (mJ/cm ²) per Log Reduction						Reference
			1	2	3	4	5	6	
MS2 DSM 5694 (Phage)	<i>E. coli</i> NCIB 9481	N/A	4	16	38	68	110		Wiedenmann et al. 1993
MS2 ATCC 15977-B1 (Phage)	<i>E. coli</i> ATCC 15977-B1	LP	15.9	34	52	71	90	109	Wilson et al. 1992
MS2 NCIMB 10108 (Phage)	<i>Salmonella typhimurium</i> WG49	N/A	12.1	30.1					Tree et al. 1997
MS2 (Phage)	<i>E. coli</i> K-12 Hfr	LP	21	36					Sommer et al. 1998
MS2 (Phage)	<i>E. coli</i> CR63	N/A	16.9	33.8					Rauth 1965
MS2 (Phage)	<i>E. coli</i> 15977	N/A	13.4	28.6	44.8	61.9	80.1		Meng and Gerba 1996
MS2 (Phage)	<i>E. coli</i> C3000	N/A	35						Battigelli et al. 1993
MS2 (Phage)	<i>E. coli</i> ATCC 15597	N/A	19	40	61				Oppenheimer et al. 1993
MS2 (Phage)	<i>E. coli</i> C3000	LP	20	42	69	92			Batch et al. 2004
MS2 (Phage)	<i>E. coli</i> ATCC 15597	LP	20	42	70	98	133		Lazarova and Savoye 2004
MS2 (Phage)	<i>E. coli</i> ATCC 15977	LP	20	50	85	120			Thurston-Enriquez et al., 2003
MS2 (Phage)	<i>E. coli</i> HS(pFamp)R	LP		45	75	100	125	155	Thompson et al. 2003
MS2 (Phage)	<i>E. coli</i> C3000	LP	20	42	68	90			Linden et al. 2002a
MS2 (Phage)	<i>E. coli</i> K-12	LP	18.5	36	55				Sommer et al. 2001
MS2 (Phage)	<i>E. coli</i> NCIMB 9481	N/A	14						Tree et al. 2005
PHI X 174 (Phage)	<i>E. coli</i> WG5	LP	2.2	5.3	7.3	10.5			Sommer et al. 1998
PHI X 174 (Phage)	<i>E. coli</i> C3000	N/A	2.1	4.2	6.4	8.5	10.6	12.7	Battigelli et al. 1993
PHI X 174 (Phage)	<i>E. coli</i> ATCC15597	N/A	4	8	12				Oppenheimer et al. 1993
PHI X 174 (Phage)	<i>E. coli</i> WG 5	LP	3	5	7.5	10	12.5	15	Sommer et al. 2001
PHI X 174 (Phage)	<i>E. coli</i> ATCC 13706	LP	2	3.5	5	7			Giese and Darby 2000
Staphylococcus aureus phage A 994 (Phage)	<i>Staphylococcus aureus</i> 994	LP	8	17	25	36	47		Sommer et al. 1989
Calicivirus canine	MDCK cell line	LP	7	15	22	30	36		Husman et al. 2004
Calicivirus feline	CRFK cell line	LP	7	16	25				Husman et al. 2004
Calicivirus feline	CRFK cell line	N/A	4	9	14				Tree et al. 2005
Calicivirus feline	CRFK cell line	LP	5	15	23	30	39		Thurston-Enriquez et al. 2003
Adenovirus type 2	A549 cell line	LP	20	45	80	110			Shin et al. 2005
Adenovirus type 2	Human lung cell line	LP	35	55	75	100			Ballester and Malley 2004
Adenovirus type 2	PLC / PRF / 5 cell line	LP	40	78	119	160	195	235	Gerba et al. 2002
Adenovirus type 15	A549 cell line (ATCC CCL-185)	LP	40	80	122	165	210		Thompson et al. 2003
Adenovirus type 40	PLC / PRF / 5 cell line	LP	55	105	155				Thurston-Enriquez et al. 2003
Adenovirus type 40	PLC / PRF / 5 cell line	LP	30	ND	ND	124			Meng and Gerba 1996
Adenovirus type 41	PLC / PRF / 5 cell line	LP	23.6	ND	ND	111.8			Meng and Gerba 1996
Poliovirus Type 1 ATCC Mahoney	N/A	N/A	6	14	23	30			Harris et al. 1987
Poliovirus Type 1 LSc2ab ()	MA104 cell	N/A	5.6	11	16.5	21.5			Chang et al. 1985

Table 4. (continued)

Virus	Host	Lamp Type	UV Dose (Fluence) (mJ/cm ²) per Log Reduction						Reference
			1	2	3	4	5	6	
Poliovirus Type 1 LSc2ab	BGM cell	LP	5.7	11	17.6	23.3	32	41	Wilson et al. 1992
Poliovirus 1	BGM cell line	N/A	5	11	18	27			Tree et al. 2005
Poliovirus 1	CaCo2 cell-line (ATCC HTB37)	LP	7	17	28	37			Thompson et al. 2003
Poliovirus 1	BGM cell line	LP	8	15.5	23	31			Gerba et al. 2002
Poliovirus Type Mahoney	Monkey kidney cell line Vero	LP	3	7	14	40			Sommer et al. 1989
Coxsackievirus B5	Buffalo Green Monkey cell line	N/A	6.9	13.7	20.6				Battigelli et al. 1993
Coxsackievirus B3	BGM cell line	LP	8	16	24.5	32.5			Gerba et al. 2002
Coxsackievirus B5	BGM cell line	LP	9.5	18	27	36			Gerba et al. 2002
Reovirus-3	Mouse L-60	N/A	11.2	22.4					Rauth 1965
Reovirus Type 1 Lang strain	N/A	N/A	16	36					Harris et al. 1987
Rotavirus SA-11	Monkey kidney cell line MA 104	LP	8	15	27	38			Sommer et al. 1989
Rotavirus SA-11	MA-104 cell line	N/A	7.6	15.3	23				Battigelli et al. 1993
Rotavirus SA-11	MA-104 cell line	N/A	7.1	14.8	25				Chang et al. 1985
Rotavirus SA-11	MA-104 cell line	LP	9.1	19	26	36	48		Wilson et al. 1992
Rotavirus	MA104 cells	LP	20	80	140	200			Caballero et al. 2004
Hepatitis A HM175	FRhK-4 cell	LP	5.1	13.7	22	29.6			Wilson et al. 1992
Hepatitis A	HAV/HFS/GBM	N/A	5.5	9.8	15	21			Wiedenmann et al. 1993
Hepatitis A HM175	FRhK-4 cell	N/A	4.1	8.2	12.3	16.4			Battigelli et al. 1993
Echovirus I	BGM cell line	LP	8	16.5	25	33			Gerba et al. 2002
Echovirus II	BGM cell line	LP	7	14	20.5	28			Gerba et al. 2002

REFERENCES

- Amoah, K., Craik, S., Smith, D.W. and Belosevic, M. 2005. Inactivation of *Cryptosporidium* oocysts and *Giardia* cysts by ultraviolet light in the presence of natural particulate matter, *AQUA, J. Wat. Supply* 54(3): 165-178.
- Ballester, N.A. and Malley, J.P. 2004. Sequential disinfection of adenovirus type 2 with UV-chlorine-chloramine, *J. Amer. Wat. Works Assoc.*, 96(10): 97-102.
- Batch, L.F., Schulz, C.R. and Linden, K.G. 2004. Evaluating water quality effects on UV disinfection of MS2 coliphage, *J. Amer. Wat. Works Assoc.*, 96(7): 75-87.
- Battigelli, D.A., Sobsey, M.D. and Lobe, D.C. 1993. The inactivation of Hepatitis A virus and other model viruses by UV irradiation, *Wat. Sci. Tech.*, 27(3-4): 339-342.
- Belosevic, M., Craik, S.A., Stafford, J.L. Neumann, N.E., Kruithof, J. and Smith, D.W. 2001. Studies on the resistance/reaction of *Giardia muris* cysts and *C. parvum* oocysts exposed to medium-pressure ultraviolet radiation, *FEMS Microbiol. Lett.*, 204(1): 197-204.
- Bolton J.R. and Linden, K.G. 2003. Standardization of methods for fluence (UV Dose) determination in bench-scale UV experiments. *J. Environ. Eng.* 129(3): 209-216.
- Bukhari, Z., Abrams, F. and LeChevallier, M. 2004. Using ultraviolet light for disinfection of finished water, *Water Sci. Tech.*, 50(1): 173-178.
- Caballero, S., Abad, F.X., Loisy, F., Le Guyader, F.S., Cohen, J., Pinto, R.M. and Bosch, A. 2004. Rotavirus virus-like particles as surrogates in environmental persistence and inactivation studies, *Appl. Env. Microbiol.* 70(7): 3904-3909.
- Campbell, A.T. and Wallis, P. 2002. The effect of UV irradiation on human-derived *Giardia lamblia* cysts, *Wat. Res.*, 36(4): 963- 969.
- Carlson, D.A., Seabloom, R.W., DeWalle, F.B., Wetzler, T.F., Engeset, J., Butler, R., Wangsuphachart, S. and Wang, S. 1985. Ultraviolet disinfection of water for small water supplies. US EPA Report No. EPA/600/S2-85/092.
- Chang, J.C.H., Osoff, S.F., Lobe, D.C., Dorfman, M.H., Dumais, C.M., Qualls, R.G. and Johnson, J.D. 1985. UV inactivation of pathogenic and indicator microorganisms, *Appl. Environ. Microbiol.*, 49(6): 1361-1365.

- Clancy, J.L., Bukhari, Z., Hargy, T.M., Bolton, J.R., Dussert, B.W. and Marshall, M.M. 2000. Using UV to inactivate *Cryptosporidium* – Even extremely low dosages of ultraviolet light can be highly effective for inactivating *Cryptosporidium* oocysts, J. Amer. Wat. Works Assoc., 92(9): 97-104.
- Clancy, J.L., Marshall, M.M., Hargy, T.M. and Korich, D.G. 2004. Susceptibility of five strains of *Cryptosporidium parvum* oocysts to UV light, J. Amer. Wat. Works Assoc., 96(3), 84-93.
- Craik, S.A., Finch, G.R., Bolton, J.R. and Belosevic, M. 2000. Inactivation of *Giardia muris* cysts using medium-pressure ultraviolet radiation in filtered water, Wat. Res., 34(18): 4325-4332.
- Craik, S.A., Weldon, D., Finch, G.R., Bolton, J.R. and Belosevic, M. 2001. Inactivation of *Cryptosporidium parvum* oocysts using medium- and low-pressure ultraviolet radiation, Wat. Res., 35(6): 1387-1398.
- Gerba, C.P., Gramos, D.M. and Nwachuku, N. 2002. Comparative inactivation of enteroviruses and adenovirus 2 by UV light, Appl. Environ. Microbiol., 68(10): 5167-5169.
- Giese, N. and Darby, J. 2000. Sensitivity of microorganisms to different wavelengths of UV light: implications on modeling of medium pressure UV systems, Wat. Res., 34(16): 4007-4013.
- Harris, G.D., Adams, V.D., Sorensen, D.L. and Curtis, M.S. 1987. Ultraviolet inactivation of selected bacteria and viruses with photoreactivation of the bacteria, Wat. Res., 21(6): 687-692.
- Hayes, S.L., Rice, E.W., Ware, M.W. and Schaefer III, F.W. 2003. Low pressure ultraviolet studies for inactivation of *Giardia muris* cysts, J. Appl. Microbiol., 94(1): 54-59.
- Hijnen, W.A.M., Beerendonk, E.F. and Medema, G.J. 2006. Inactivation credit of UV radiation for viruses, bacteria and protozoan (oo)cysts in water; a review, Wat. Res., 40(1): 3-22.
- Hoyer, O. 1998. Testing performance and monitoring of UV systems for drinking water disinfection, Wat. Supply, 16(1-2): 424-429.
- Huffman, D.E., Gennaccaro, A., Rose, J.B. and Dussert, B.W. 2002. Low- and medium-pressure UV inactivation of microsporidia *Encephalitozoon intestinalis*, Wat. Res., 36(12): 3161-3164.
- Husman, A.M.D., Bijkerk, P., Lodder, W., Van den Berg, H., Pribil, W., Cabaj, A., Gehringer, P., Sommer, R. and Duizer, E. 2004. Calicivirus inactivation by nonionizing 253.7-nanometer-wavelength (UV) and ionizing (Gamma) radiation, Appl. Environ. Microbiol., 70(9): 5089-5093.
- Johnson, A.M., Linden, K., Ciociola, K.M., De Leon, R., Widmer, G. and Rochelle, P.A. 2005. UV inactivation of *Cryptosporidium hominis* as measured in cell culture, Appl. Environ. Microbiol., 71(5): 2800-2802.
- Joux, F., Jeffrey, W.H., Lebaron, P. and Mitchell, D. L. 1999. Marine bacterial isolates display diverse responses to UV-B radiation, Appl. Environ. Microbiol., 65(9): 3820-3827.
- Karanis, P., Maier, W.A., Seitz, H.M. and Schoenen, D. 1992. UV sensitivity of protozoan parasites, Aqua, 41: 95-100.
- Lazarova, V. and Savoye, P. 2004. Technical and sanitary aspect of wastewater disinfection by ultraviolet irradiation for landscape irrigation, Wat. Sci. Technol., 50(2): 203-209.
- Liltved, H. and Landfald, B. 1996. Influence of liquid holding recovery and photoreactivation on survival of ultraviolet-irradiated fish pathogenic bacteria, Wat. Res., 30(5): 1109-1114.
- Linden, K.G., Batch, L. and Schulz, C. 2002a. UV disinfection of filtered water supplies: water quality impacts on MS2 dose-response curves, Proceedings Amer. Wat. Works Assoc. Annu. Conf., Amer. Wat. Works Assoc., Denver, CO.
- Linden, K.G., Shin, G.-A., Faubert, G., Cairns, W. and Sobsey, M.D. 2002b. UV disinfection of *Giardia lamblia* cysts in water, Environ. Sci. Technol., 36(11): 2519-2522.
- Mamane-Gravetz, H. and Linden, K.G. 2004. UV disinfection of indigenous aerobic spores: Implications for UV reactor validation in unfiltered waters, Wat. Res., 38(12): 2898-2906.
- Marshall, M.M., Hayes, S., Moffett, J., Sterling, C.R. and Nicholson, W.L. 2003. Comparison of UV inactivation of three *Encephalitozoon* species with that of spores of two DNA repair-deficient *Bacillus subtilis* biosimetry strains, Appl. Environ. Microbiol., 69(1): 683-685.
- Martin, E.L., Reinhardt, R.L., Baum, L.L., Becker, M.R., Shaffer, J.J. and Kokjohn, T.A. 2000. The effects of ultraviolet radiation on the moderate halophile *Halomonas elongata* and the extreme halophile *Halobacterium salinarum*, Can. J. Microbiol., 46(2): 180-187.
- Maya, C., Beltran, N., Jimenez, B. and Bonilla, P. 2003. Evaluation of the UV disinfection process in bacteria and amphizoic amoebae inactivation, Wat. Sci. Technol.: Wat. Supply, 3(4): 285-291.
- Meng, Q.S. and Gerba, C.P. 1996. Comparative inactivation of enteric adenoviruses, poliovirus and coliphages by ultraviolet irradiation, Wat. Res., 30(11): 2665-2668.
- Mofidi, A.A., Meyer, E.A., Wallis, P.M., Chou, C.I., Meyer, B.P., Ramalingam, S. and Coffey, B.M. 2002. The effect of UV light on the inactivation of *Giardia lamblia* and *Giardia muris* cysts as determined by animal infectivity assay, Wat. Res., 36(8): 2098-2108.
- Morita, S., Namikoshi, A., Hirata, T., Oguma, K., Katayama, H., Ohgaki, S., Motoyama, N. and Fujiwara, M. 2002. Efficacy of UV irradiation in inactivating *C. parvum* oocysts, Appl. Environ. Microbiol., 68(11): 5387-5393.
- Nieuwstad, T.J. and Havelaar, A.H. 1994. The kinetics of batch ultraviolet inactivation of bacteriophage MS2 and microbiological calibration of an ultraviolet pilot plant, J. Environ. Sci. Health, A29(9): 1993-2007.
- Oguma, K., Katayama, H. and Ohgaki, S. 2002. Photoreactivation of *E. coli* after low- or medium-pressure UV disinfection determined by an endonuclease sensitive site assay, Appl. Environ. Microbiol., 68(12), 6029-6035.
- Oguma, K., Katayama, H. and Ohgaki, S. 2004. Photoreactivation of *Legionella pneumophila* after inactivation by low- or medium-pressure ultraviolet lamp, Wat. Res., 38(11): 2757-2763.
- Oppenheimer, J.A., Hoagland, J.E., Laine, J.-M., Jacangelo, J.G. and Bhamrah, A. 1993. Microbial inactivation and characterization of toxicity and by-products occurring in reclaimed wastewater disinfected with UV radiation, Conf. on Planning, Design and Operation of Effluent Disinfection Systems, Whippany, NJ, May 23-25, 1993, Wat. Environ. Fed., Alexandria, VA

- Otaki, M., Okuda, A., Tajima, K., Iwasaki, T., Kinoshita, S. and Ohgaki, S. 2003. Inactivation differences of microorganisms by low pressure UV and pulsed xenon lamps, *Wat. Sci. Technol.*, 47(3): 185-190.
- Rauth, A.M. 1965. The physical state of viral nucleic acid and the sensitivity of viruses to ultraviolet light, *Biophys. J.*, 5: 257-273.
- Rice, E.W. and Hoff, J.C. 1981. Inactivation of *Giardia lamblia* cysts by ultraviolet irradiation, *Appl. Environ. Microbiol.*, 42(3): 546-547.
- Shin, G.-A., Linden, K.G. and Sobsey, M.D. 2000. Comparative inactivation of *Cryptosporidium parvum* oocysts and coliphage MS2 by monochromatic UV radiation, *Proceedings of Disinfection 2000: Disinfection of Wastes in the New Millennium*, New Orleans, Water Environment Federation, Alexandria, VA.
- Shin, G.-A., Linden, K.G., Arrowood, M.J. and Sobsey, M.D. 2001. Low-pressure UV inactivation and DNA repair potential of *C. parvum* oocysts, *Appl. Environ. Microbiol.*, 67(7): 3029-3032.
- Shin, G.A., Linden, K.G. and Sobsey, M.D. 2005. Low pressure ultraviolet inactivation of pathogenic enteric viruses and bacteriophages, *J. Environ. Engr. Sci.*, 4: S7-S11.
- Sommer, R., Weber, G., Cabaj, A., Wekerle, J., Keck, G., and Schaubberger, G. 1989. UV inactivation of microorganisms in water. *Zbl. Hyg.* 189: 214-224.
- Sommer, R., Haider, T., Cabaj, A., Pribil, W. and Lhotsky, M. 1998. Time dose reciprocity in UV disinfection of water, *Water Sci. Technol.*, 38(12): 145-150.
- Sommer, R., Cabaj, A., Sandu, T. and Lhotsky, M. 1999. Measurement of UV radiation using suspensions of microorganisms, *J. Photochem. Photobiol.*, 53(1-3): 1-5.
- Sommer, R., Lhotsky, M., Haider, T. and Cabaj, A. 2000. UV inactivation, liquid-holding recovery, and photoreactivation of *E. coli* O157 and other pathogenic *E. coli* strains in water, *J. Food Protection*, 63(8): 1015-1020.
- Sommer, R., Pribil, W., Appelt, S., Gehringer, P., Eschweiler, H., Leth, H., Cabaj, A. and Haider, T. 2001. Inactivation of bacteriophages in water by means of non-ionizing (UV-253.7 nm) and ionizing (gamma) radiation: A comparative approach, *Wat. Res.*, 35(13): 3109- 3116.
- Thurston-Enriquez, J.A., Haas, C.N., Jacangelo, J., Riley, K. and Gerba, C.P. 2003. Inactivation of feline calicivirus and adenovirus type 40 by UV radiation, *Appl. Environ. Microbiol.*, 69(1): 577-582.
- Thompson, S.S., Jackson, J.L., Suva-Castillo, M., Yanko, W.A., Jack, Z.E., Kuo, J., Chen, C.L., Williams, F.P. and Schnurr, D.P. 2003. Detection of infectious human adenoviruses in tertiary-treated and ultraviolet-disinfected wastewater, *Wat. Environ. Res.*, 75(2): 163-170.
- Tosa, K. and Hirata, T. 1998. HRWM-39: Photoreactivation of *Salmonella* following UV disinfection, *IAWQ 19th Biennial International Conference*, Vol. 10, Health-Related Water Microbiology.
- Tosa, K. and Hirata, T. 1999. Photoreactivation of enterohemorrhagic *E. coli* following UV disinfection, *Wat. Res.*, 33(2): 361-366.
- Tree, J.A., Adams, M.R. and Lees, D.N. 1997. Virus inactivation during disinfection of wastewater by chlorination and UV irradiation and the efficacy of F+ bacteriophage as a 'viral indicator', *Wat. Sci. Technol.*, 35(11-12): 227-232.
- Tree, J.A., Adams, M.R. and Lees, D.N. 2005. Disinfection of feline calicivirus (a surrogate for Norovirus) in wastewaters, *J. Appl. Microbiol.*, 98: 155-162.
- Wiedenmann, A., Fischer, B., Straub, U., Wang, C.-H., Flehmig, B. and Schoenen, D. 1993. Disinfection of Hepatitis A virus and MS-2 coliphage in water by ultraviolet irradiation: Comparison of UV-susceptibility, *Wat. Sci. Tech.*, 27(3-4): 335-338.
- Wilson, B.R., Roessler, P.F., Van Dellen, E., Abbaszadegan, M. and Gerba, C.P. 1992. Coliphage MS-2 as a UV water disinfection efficacy test surrogate for bacterial and viral pathogens, *Proceedings, Water Quality Technology Conference*, Nov 15-19, 1992, Toronto, Canada, pp. 219-235, Amer. Wat. Works Assoc., Denver, CO.
- Wu, Y., Clevenger, T. and Deng, B. 2005. Impacts of goethite particles on UV disinfection of drinking water, *Appl. Environ. Microbiol.*, 71(7): 4140-4143.
- Yaun, B.R., Sumner, S.S., Eifert, J.D. and Marcy, J.E. 2003. Response of *Salmonella* and *E. coli* O157:H7 to UV energy, *J. Food Protection*, 66(6): 1071-1073.
- Zimmer, J.L. and Slawson, R.M. 2002. Potential repair of *E. coli* DNA following exposure to UV radiation from both medium- and low-pressure UV sources used in drinking water treatment, *Appl. Environ. Microbiol.*, 68(7): 3293-3299.
- Zimmer, J.L., Slawson, R.M. and Huck, P.M. 2003. Inactivation and potential repair of *C. parvum* following low- and medium-pressure ultraviolet irradiation, *Wat. Res.*, 37(14): 3517-3523.